#### Bioresource Technology 116 (2012) 179-183

Contents lists available at SciVerse ScienceDirect

# **Bioresource Technology**

journal homepage: www.elsevier.com/locate/biortech

# Effect of carbon and nitrogen sources on photo-fermentative H<sub>2</sub> production associated with nitrogenase, uptake hydrogenase activity, and PHB accumulation in *Rhodobacter sphaeroides* KD131

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## ARTICLE INFO

Article history: Received 18 January 2012 Received in revised form 3 April 2012 Accepted 5 April 2012 Available online 13 April 2012

Keywords: Biohydrogen Photo-fermentation Nitrogenase Uptake hydrogenase Poly-β-hydroxybutyrate

#### ABSTRACT

During photo-fermentative  $H_2$  production, the effects of carbon and nitrogen sources on nitrogenase and hydrogenase activity, poly- $\beta$ -hydroxybutyrate accumulation were investigated. In succinate/ammonium sulfate medium,  $H_2$  was not detected for the first 6 h because high ammonium concentration considerably reduced the nitrogenase activity to below 5 nmol/g-dcw/h. After 24 h, 99% of the ammonium was consumed, and the nitrogenase activity increased to 296 nmol/g-dcw/h, accelerating  $H_2$  production. In contrast, the ammonium in succinate/glutamate medium was much less, which led to rapid  $H_2$  production in the beginning. However,  $H_2$  evolution was repressed over time by increased ammonium. In the presence of  $H_2$ , hydrogenase activity increased with time regardless of the nitrogen source, and consequently,  $H_2$  production was reduced. Compared with succinate,  $H_2$  production in acetate media was severely limited due to increased pH over 9. During extended cultivation, the PHB accumulated in acetate media.

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## 1. Introduction

Hydrogen  $(H_2)$  is regarded as an environmentally clean fuel to overcome the adverse effect of hydrocarbon fuels where carbon dioxide and other pollutants are released to the atmosphere. Since H<sub>2</sub> carries high energy per unit mass and produces only water vapor at the end point of use, extensive studies have been done on the hydrogen energy system, including production, storage, use, and safety issues. However, most hydrogen fuels currently are produced by reforming of fossil fuels using physicochemical methods which are usually expensive and energy intensive. Thus, biological H<sub>2</sub> production from renewable energy sources, such as biomass, is considered the most environmental friendly technology. It has advantages of reducing organic waste and generating clean energy, i.e. hydrogen. In particular, phototrophic fermentation using purple non-sulfur (PNS) bacteria is of great interest due to high theoretical H<sub>2</sub> yield from various volatile organic acids (Chen et al., 2011).

During photo-fermentation,  $H_2$  is produced by the interactions of several metabolic pathways. PNS bacteria evolve  $H_2$  through catalysis of nitrogenase with the consumption of adenosine

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triphosphate (ATP) and free electrons originating from light as follows:

$$2H^+ + 2e^- + 4ATP \rightarrow H_2 + 4ADP + 4P_i \tag{1}$$

The activity of nitrogenase is crucial to H<sub>2</sub> production by photosynthetic bacteria, and it is strongly inhibited by high ammonia concentration (Koku et al., 2002). Thus, the concentration and type of nitrogen source is critical to the performance of phototrophic H<sub>2</sub> production. The substrate is also a key factor affecting cell growth and H<sub>2</sub> production (Khatipov et al., 1998; Fang et al., 2005). In the aspect of substrate conversion efficiency, cell growth and alternative biosynthesis are electron sinkers to reduce the portion of the substrate available for H<sub>2</sub> production. Among metabolite alternatives to H<sub>2</sub>, the bacterial storage compound, poly-β-hydroxybutyrate (PHB) is closely associated with H<sub>2</sub> production (Hustede et al., 1993; Kim et al., 2006). The polymer, which serves as a reserve of carbon and energy, is accumulated in cytoplasm under unbalanced growth conditions. The amount of PHB accumulated in photosynthetic bacteria depends on physiological conditions, such as pH and the type of substrate (Hustede et al., 1993; Khatipov et al., 1998). The total H<sub>2</sub> production is also limited by uptake hydrogenase activity. This enzyme catalyzes the conversion of molecular H<sub>2</sub> to protons and electrons to reduce a relatively high potential electron acceptor, and it consequently decreases the efficiency of H<sub>2</sub> production (Kars et al., 2008).



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