



Factors affecting cellular lipid extraction from marine microalgae

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HIGHLIGHTS

- ▶ Investigate key factors affecting extraction efficiency and lipid characteristics.
- ▶ FFA was increased 3 times in extracted lipid for microalgae dried by solar.
- ▶ Chlorinated solvent systems resulted in higher extraction efficiencies than other solvent.
- ▶ Hexane had a poor extraction efficiency, but improved when polar solvents were added.
- ▶ The moisture content affected both lipid extraction efficiency and FFA content.

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ABSTRACT

Factors affecting intracellular lipid extraction from marine microalgae were investigated using various techniques. The biomass drying method and moisture content, and the solvent extraction system were the factors studied. Lipid was analytically classified into three categories i.e. neutral lipid, free fatty acid (FFA) and polar lipid using solid-phase extraction. Biomass drying methods (freeze-, oven- and solar drying) did not affect lipid yield, but the FFA content of the lipid was three times higher for solar dried biomass. Of the various lipid extraction methods tested, including sonication, homogenization, accelerated solvent extraction (ASE) and Soxhlet extraction, sonication was the least efficient compared to other methods when a partially miscible solvent system i.e. hexane–methanol was used. Chloroform–methanol solvent system had maximum lipid extraction efficiency (33%). A biomass moisture content up to 5% had no impact on lipid extraction efficiency, but higher moisture contents reduced lipid extraction and increased the FFA fraction.

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1. Introduction

Microalgae are currently subject to worldwide investigation as a promising sustainable and renewable energy source to meet future energy demand for liquid transportation fuels. Using microalgae to tap solar energy via photosynthesis is not a new concept; where an extensive study was conducted under the United States Aquatic Species Program [1]. Moreover, microalgae production represents a potential solution for the mitigation of climate change since 1 ton of algae results in the fixation of 1.83 tons of atmospheric CO₂ [2]. The yield of lipid–oil feedstock from microalgae has been estimated at 123 m³/hectare which is up to 20 times higher when compared to palm oil [2]. Chisti [2] estimated that if all US petroleum-based transport fuels were to be replaced by biodiesel, then palm oil plantation would require 61% of the total

cultivable land of the continental United States; whereas microalgae would need only 3%, assuming the microalgae contained 15% lipid oil as biomass dry weight.

Processing of harvested microalgal biomass to extract lipid–oil can be an energy intensive process as it requires separation and dewatering of microalgae, biomass drying and the subsequent extraction and purification of lipid–oil. The crude extract contains not only lipid–oils, but also carbohydrates, proteins and pigments. The lipids can be further classified into neutral lipids, free fatty acids, and polar lipids which include galacto- and phospholipids [3]. The lipid composition of various types of microalgae has been studied by several researchers, for example diatoms [3], *Nannochloropsis* [4] and *Phaeodactylum tricornutum* [5]. Lee et al. [6] studied various lipid extraction methods and solvent systems and compared relative extraction efficiencies. It was found that cell disruption with a micro-bead beater followed by extraction with chloroform:methanol (2:1 v/v) was an effective method for lipid extraction [7]. Addition of concentrated hydrochloric acid to a chloroform:methanol solvent system increased the total lipids extracted, especially phospholipids, in microalgae i.e. *Botryococcus*

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